

Biodegradation

How UV photolysis accelerates the biodegradation and mineralization of sulfadiazine (SD) --Manuscript Draft--

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Abstract:	Sulfadiazine (SD), one of broad-spectrum antibiotics, exhibits limited biodegradation in wastewater treatment due to its chemical structure, which requires initial mono-oxygenation reactions to initiate its biodegradation. Intimately coupling UV photolysis with biodegradation, realized with the internal loop photobiodegradation reactor (ILPBR), accelerated SD biodegradation and mineralization by 40% and 71%, respectively. The main organic products from photolysis were 2-aminopyrimidine (2-AP), p-aminobenzenesulfonic acid (ABS), and aniline (An), and an SD-photolysis pathway could be identified using C, N, and S balances. Adding An or ABS (but not 2-AP) into the SD solution during biodegradation experiments (no UV photolysis) gave SD removal and mineralization rates similar to intimately coupled photolysis and biodegradation. An SD biodegradation pathway, based on a diverse set of the experimental results, explains how the mineralization of ABS provided internal electron carriers that accelerated the initial mono-oxygenation reactions of SD biodegradation. Thus, multiple lines of evidence support that the mechanism by which intimately coupled photolysis and biodegradation accelerated SD removal and mineralization was through producing co-substrates whose oxidation produced electron equivalents that stimulated the initial mono-oxygenation reactions for SD biodegradation.
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How UV photolysis accelerates the biodegradation and mineralization of sulfadiazine (SD)

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Abstract:

Sulfadiazine (SD), one of broad-spectrum antibiotics, exhibits limited biodegradation in wastewater treatment due to its chemical structure, which requires initial mono-oxygenation reactions to initiate its biodegradation. Intimately coupling UV photolysis with biodegradation, realized with the internal loop photobiodegradation reactor (ILPBR), accelerated SD biodegradation and mineralization by 40% and 71%, respectively. The main organic products from photolysis were 2-aminopyrimidine (2-AP), *p*-aminobenzenesulfonic acid (ABS), and aniline (An), and an SD-photolysis pathway could be identified using C, N, and S balances. Adding An or ABS (but not 2-AP) into the SD solution during biodegradation experiments (no UV photolysis) gave SD removal and mineralization rates similar to intimately coupled photolysis and biodegradation. An SD biodegradation pathway, based on a diverse set of the experimental results, explains how the mineralization of ABS provided internal electron carriers that accelerated the initial mono-oxygenation reactions of SD biodegradation. Thus, multiple lines of evidence support that the mechanism by which intimately coupled photolysis and biodegradation accelerated SD removal and mineralization was through producing co-substrates whose oxidation produced electron equivalents that stimulated the initial mono-oxygenation reactions for SD biodegradation.

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1. Introduction

Sulfonamides (SAs) are broad-spectrum antibiotics widely used for prevention and treatment of animal diseases and promotion of livestock growth (Zhao et al., 2010; Liu and Wong, 2013). SAs are present in wastewaters involved with livestock production (Žegura et al., 2009), and this poses a health risk to human being when the SAs enter supplies of potable water (Kim and Tanaka, 2009; Periša M et al., 2013). Among the SAs, sulfadiazine (SD, $C_{10}H_{10}N_4O_2S$) is an important representative.

Many physical, chemical, and biological methods may be employed to reduce or remove SAs from wastewaters (Garcia et al., 2012). Among them, advanced oxidation processes (AOPs) show promise for SD degradation: e.g., ultrasound + hydrogen peroxide (Mohajerani et al., 2012), Fenton's reaction (Neafsey et al., 2009), ozonation (Zhou et al., 2011), TiO_2 photocatalytic oxidation (Baran et al., 2009), and UV photolysis (Yan et al., 2012). AOPs alone are not practical to mineralize SD, but they can modify the SD structure in ways that make it more readily removed by another process, such as biodegradation. Therefore, a good strategy is to couple an AOP with biodegradation for complete degradation of SAs. The AOP produces partial-breakdown products that accelerate the biodegradation and mineralization by being more readily biodegradable than the original recalcitrant compound (Zhang et al., 2012a), having less toxicity (Zhang et al., 2012b), or being oxidized to produce intracellular electron carriers that accelerate initial reactions (such as mono-oxygenations) for the biodegradation of the original compound (Zhang et al., 2014). Thus, coupling an AOP to biodegradation can increase the removal rate of the original compound by its direct action on the molecule and by its indirect effects.

A promising means to couple an AOP with biodegradation is called intimate coupling, in which the AOP produces transformation products that are rapidly consumed by bacteria present in the same reactor (Marsolek et al., 2008; Li et al., 2012; Zhang et al., 2014). Intimate coupling overcomes problems that often plague a sequentially coupled system: Too much photolysis increases costs and can generate products that are inhibitory or recalcitrant. Thus, intimate coupling allows intermediates to be biodegraded as soon they are formed.

Here, we focus on the benefits of intimately coupling UV photolysis with biodegradation for SD. We hypothesize that UV photolysis generates a series of biodegradable intermediates, and bacterial oxidation of the intermediates produces intracellular electron carriers for initial mono-oxygenation reactions needed to biodegrade SD (Sukul et al., 2008; Lafaille et al., 2014). Similar accelerations have been seen for biodegradation of 2,4-dichlorophenol (Dahlen and Rittmann, 2000), toluene and *p*-xylene (Yu et al., 2001), and pyridine (Zhang et al., 2014). Having a more complex structure, SD will release more and diverse photolysis products, and their ability to be oxidized to supply intracellular electron carriers is unknown.

In this work, we used an internal loop photolytic biofilm reactor (ILPBR) (Zhang et al. 2012b) to realize intimate coupling of UV photolysis with biodegradation for SD. We document that intimate coupling gave faster degradation and mineralization kinetics than photolysis alone or biodegradation alone. Just as importantly, we elaborate the mechanism by which intimately coupled UV photolysis accelerates SD biodegradation.

2. Materials and methods

2.1. Chemicals

All chemicals were purchased through Sinopharm Chemical Reagent Co., Ltd in Shanghai, China. SD and *p*-aminobenzenesulfonic acid were from Sigma Co. Ltd. (USA); acetonitrile from Tedia (USA); methanol from Anaqua Chemicals Supply Co. Ltd. (USA); and 2-aminopyrimidine, aniline, acetic acid, and all inorganic salts from Sinopharm Chemical Reagent Co., Ltd in Shanghai (China).

2.2. Inorganic salt, trace-element, and buffer stock solutions

The inorganic salt solution was prepared by diluting into 250 mL of deionized water 4.775 g NH_4Cl and 1.097g KH_2PO_4 to give a solution with 5 g N/L and 1 g P/L. The trace-element solution was (g/L): $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$ 1.5, $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$ 0.024, $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$ 0.19, $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ 0.002, $\text{MnSO}_4 \cdot 7\text{H}_2\text{O}$ 0.1, $\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$ 0.024, ZnCl_2 0.07, and H_3BO_3 0.006. The buffer solution was (g/L): KH_2PO_4 8.5, K_2HPO_4 21.75, $\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$ 33.4, and NH_4Cl 1.7. These stock solutions were stored in a refrigerator at 4°C.

2.3. Preparation of SD solutions

1 g SD was diluted in 800 mL deionized water, and the pH was adjusted to 10 with 0.1 M NaOH. Once the SD was fully dissolved, more deionized water was added to give 1000 mL, which gave an SD stock solution of 1000 mg/L. It was stored in a refrigerator at 4°C. During an experiment, the stock solution was diluted with deionized water to give an SD concentration in the

range of 5 to 100 mg/L. All solutions were then adjusted to pH 7 by addition of the buffer solution, and 1 mL each of the inorganic salt and trace element solutions were added for each 1 L of solution used for biodegradation experiments.

2.4. Internal loop photo-biodegradation reactor (ILPBR)

An internal loop photolytic biofilm reactor (ILPBR) was made of quartz glass with a 45-mL working volume ([Zhang et al., 2012b](#)). A glass baffle was installed in the center of the ILPBR to create photolysis and biodegradation zones within one reactor. The biofilm carriers in the biodegradation zone were hydroformylated fibers produced by an aldehyde reaction; they were soft plastic fibers having a rough outer surface ideal for biofilm attachment. An air-lift pump circulated water between the photolysis zone and the biodegradation zone at a rate of 80 mL/min. When only UV photolysis was carried out, the biofilm carriers were not present, and the UV light was on. When only biodegradation was carried out, the biofilm carriers were installed in the ILPBR, but the UV light was off. Intimately coupled UV photolysis and biodegradation had biofilm carriers installed and the UV light on.

2.5. Acclimation of SD-degrading bacteria and preparation of biofilm

Activated sludge was obtained from the underflow of a secondary clarifier at the Longhua municipal wastewater treatment plant in Shanghai. For acclimation under aerobic conditions, 100 mL sludge was added into a 500-mL graduated cylinder; fresh medium with 300 mg/L of COD was replaced every day for seven days, and the temperature was 25 – 30°C. The medium was prepared

with glucose, ammonium chloride, and potassium dihydrogen phosphate based on the mass ratio of COD:N:P = 100:5:1, and it also included 1 mL each of inorganic salt, buffer, and trace element solutions.

For each of the next 14 days, we partially replaced glucose with SD in the medium. In 5 steps, we decreased the glucose concentration from 300 mg/L to 260 mg/L, while increasing the SD concentration from zero to 20 mg/L to keep the same COD concentration (300 mg/L). During the acclimation, we replaced the medium every day, and the acclimation strategy was continued until the SD removal percentage exceeded 98% in 3 days.

To harvest SD-degrading bacteria, the acclimated sludge was settled, the supernatant was filtered through coarse filter paper (pore size 80~120 μ m), and the filtrate contained the SD-degrading bacteria. The filtrate was stored in a refrigerator at 4°C until use, which occurred within 5 days.

To obtain SD-degrading biofilm, fibers were immersed in the acclimated sludge for another 5 days. The medium containing 20 mg/L SD and 260 mg/L glucose was replaced every day until visible biofilm accumulated on the fiber.

2.6. Experiments in the ILPBR

Before SD degradation, control experiments were carried out in the ILPBR with heat-inactivated biofilm (Yan et al., 2012) and without UV illumination to investigate any adsorption of SD to the biofilm or the carrier. Then, three protocols -- photolysis alone (P), biodegradation alone (B), and simultaneous photolysis and biodegradation (P&B) -- were used for

evaluating degradation of SD in batch experiments at 25°C. During each experiment, samples were taken at time intervals to measure SD concentrations. P experiments were carried out with the UV light turned on, no biofilm present in the biodegradation zone, and the solution circulated between the photolysis zone and the biodegradation zone by airlift pumping. The UV wavelength was 254 nm, the power was 24 W, the light intensity was about 1.0 mW/cm², and the distance between the UV light and reactor was 10 cm. For the B experiments, biofilm-coated fibers were installed in the reactor, the UV light was turned off, and liquid circulation was driven by airlift aeration as in the P experiments. For the P&B experiments, biofilm was installed in the ILPBR, and the UV light was on at the same time.

We also conducted a series of experiments in which we evaluated the effects of UV-hydrolysis products on biodegradation of SD. All of the experiments were carried out with 16 mg/L SD and the B protocol, but had different additions of photolysis products. In one experiment, the products were generated by exposing a 20-mg/L SD solution to UV illumination for 0.5 hour, which decreased the SD concentration to 16 mg/L; this medium was then used for a B experiment. We also conducted experiments in which one or all three of the main photolysis products (ABS, An, and 2-AP) were added at concentrations equal to what was formed in a P experiment. The starting materials and nomenclature for each experiment are given in Table 1.

2.7. Fate of N and S in SD

In order to elaborate the SD-biodegradation pathway, we carried out special B and P&B experiments having an initial SD concentration of 10 mg/L. We measured NH₄⁺ and SO₄²⁻

concentrations in parallel to the loss of SD. This allowed us to relate the release of N and S to the loss of SD. For these experiments, we allowed the SD solutions to mix in the ILPBR for 2 min before taking the first sample. We designate this first sample as time = 0.

2.8. Analytical methods

SD and 2-AP were measured by a high performance liquid chromatograph (model: Agilent 1100, USA) equipped with a diode array detector with wavelength of 269 nm and ZORBAX SB-C18 column (5 μ m, 4.6 \times 150 mm). The mobile phase was a mixture of acetonitrile–water solution (25:75, v/v), its pH was adjusted with acetic acid to 4, and the flow rate was 1 mL/min. The column temperature was kept at 35°C, a 20- μ L sample was injected, and the detection limit was 5 ng/mL (Zhou et al., 2011). An and ABS were measured with the same ZORBAX SB-C18 column, but the wavelength was 205 nm, and the mobile phase was 0.1 M ammonium dihydrogen phosphate solution with 1 mL/min flow rate. The column temperature was kept at 25°C, 20 μ L was injected, and the detection limit was 5 μ g/L (Wang et al., 2002).

Inorganic ions were measured with ion chromatography (model: DIONEX ICS-5000, USA). For the ammonium cation, the separation column was IonPac CG12A (50 \times 4mm, S/N 022328), and the mobile phase (eluent) was 20 mM (methanesulfonic acid) MSA at 1.0 mL/min. For sulfate, nitrate, and nitrite anions, the separation column was IonPac AG18 (50 \times 4 mm), and the eluent was mixed solution of Na₂CO₃ (4.5 mM) and NaHCO₃ (0.8 mM) at a flow rate of 1.0 mL/min.

The TOC concentration was analyzed by using a TOC analyzer (Model: SHIMADZU, TOC-VCSN). The UV-light intensity was measured by an illuminometer (model BG-2254, China). Optical density (OD) was measured by spectrophotometer (model UV-2550, SHIMADZU,

Japan) with a wavelength of 600 nm. The biomass concentration as dry weight was calculated based on a calibration curve of OD versus dry weight ([von Stosch et al. 2012](#); [Wan et al. 2012](#)).

3. Results and discussions

3.1. ILPBR for SD photo-biodegradation and mineralization

Figure 1 compares the results for SD removal with protocols P, B, and P&B in the ILPBR over a 4-hour period with an initial SD concentration of 22 mg/L. The control results (no biodegradation or photolysis) also are shown and indicate minimal loss after 10 minutes. The most important finding in Figure 1 is that SD removal by P&B was faster than that by B or by P. In order to compare the kinetics for each protocol directly, we approximated the SD-removal kinetics as zero order after the first 10 min. The rates are 5.0 mg/L-h by P&B, 4.5 mg/L-h by P, and 3.7 mg/L-h by B; thus, P&B was 40% faster than B.

The corresponding TOC removal percentages, reflecting SD mineralization at the end of the 4-h B and P&B experiments, are shown in Figure 2. Protocol P did not give any TOC removal and is not shown. P&B gave 60% TOC removal, which is higher by 71% than for protocol B, which had 35% TOC removal. Thus, intimately coupled photolysis and biodegradation accelerated SD mineralization even more than was its impact on biodegradation of SD.

3.2. SD photolysis products and pathway

To investigate SD intermediates from UV photolysis, we illuminated a 20-mg/L SD solution for a 4-h P experiment. Figure 3 shows the photolysis products as SD declined from 20 mg/L to 1.1 mg/L. The dominant organic products (top panel) were 2-AP, ABS, and An. Sulfate and ammonia were released in parallel (bottom panel). As before, TOC removal was negligible and is not shown.

Following the pathway patterns observed previously for SD by Wang et al. (2010) and the mass-balance strategy of Zhang et al. (2013), we developed the SD-photolysis pathway shown in Figure 4. The C, N, and S balances that underlie the pathway are summarized in Table 2. The mass balance errors are < 1% for all elements in Table 2. ABS ($C_6H_7NO_3S$) and 2-AP ($C_4H_5N_3$) are produced by hydrolysis reaction A; An (C_6H_7N) and SO_4^{2-} are produced by another hydrolysis reaction B; 2-hydroxyhex-2,4-diendioic acid ($C_6H_6O_5$) and NH_3 are produced by set of oxidations in reaction C; and formic acid (CH_2O_2) and NH_3 are produced by oxidations and hydrolysis in reaction D. Significant are that ammonia is released from reactions C and D and that releases of aniline and sulfate require reaction B, the transformation of ABS.

3.3. Accelerating SD biodegradation by means of co-substrates

Based on the experimental results above, we hypothesized that one cause for the increase in SD removal and mineralization rates is that microbial oxidation of SD photolysis products generates electron equivalents that can be used as co-reactants for initial mono-oxygenation reactions; this is a type of co-substrate effect (Dahlen and Rittmann, 2000; Yu et al., 2001; Zhang et al., 2014). Thus, we performed B experiments with different combinations of the main photolysis products, as shown in Table 1.

The experimental results are shown in Figure 5, and zero-order-loss rates (mg SD/L-h), obtained by linear regression for results after 10 min, are indicated by the slopes of the dashed lines, which match the experimental data well. The results in the top panel of Figure 5 show that removal of SD in the synthetic SD solution with all photolysis products (SD+An+2-AP+ABS) was

almost the same as with the products produced by photolysis directly (SD(P)), and both were about 50% faster than with no photolysis products (SD(O)). The bottom panel of Figure 5 indicates that adding only 2-AP did not accelerate SD removal, but SD+An and SD+ABS gave accelerations about as strong as SD(P). These trend supports that microbial oxidation of An and ABS (but not 2-AP) provided reduced electron carriers needed as co-substrates for mono-oxygenation reactions used to initiate SD catabolism.

3.4. SD biodegradation pathway

We tracked the fate of N and S equivalents, together with SD biodegradation, in B and P&B experiments having an initial SD concentration of 10 mg/L. The results, shown in Figure 6, show that SD was degraded much faster in the P&B experiment, as expected. The most important finding is that N and S releases were nearly equal to the N and S releases predicted from full mineralization of SD. We did not measure the TOC in these experiments, but Figure 2 reports that the TOC removal was 60% for 100% removal of SD in its P&B experiment. If that pattern held true for the experiments of Figure 6, then the biodegradation pathway proceeded far enough that N and S no longer remained in any organic intermediates, even though not all of the C was oxidized to CO₂.

Based on our results (Figures 1, 2, 5, and 6) and information from the literature, we propose the SD-biodegradation pathway in Figure 7. The first and second steps (A and B) of SD biodegradation are mono-oxygenations that form dihydroxyl sulfadiazine. They require molecular oxygen (O₂) and an intracellular electron donor (represented as 2H). Step C is hydrolysis of

dihydroxyl sulfadiazine, which produces *p*-aminobenzenesulfonic acid (ABS) and 2-amino-4,6-dihydropyrimidine (Wang et al., 2010). The ABS is further hydrolyzed to generate aniline and sulfate (step D) (Takeo et al., 1998). Aniline mineralization is initiated by means of a mono-oxygenation reaction (step E) (Takeo et al., 1998; Baran et al., 2009b; Sukul et al., 2008; Wang et al., 2010; Neafsey et al., 2009) that requires O_2 and $2H$ and that inserts an $-OH$ group into the ring (Li et al., 2003) to form aminophenol (C_6H_7NO). This is followed by a hydrolytic substitution (step F) that replaces the amino group with hydroxyl group to form catechol ($C_6H_4O_2$) while generating ammonia (NH_3) simultaneously. Step G is a set of one mono-oxygenation reaction and two hydroxylation reactions that give 2-hydrohexa-2,4-diendioic acid (HDA, $C_6H_6O_5$). Step H begins mineralization of HAD to form CO_2 and H_2O , and it releases 20 electron equivalents via a series of hydroxylation and dehydrogenation reactions. Step I represents a set of hydration reactions that decompose 2-amino-4,6-dihydropyrimidine into formic acid and NH_3 . The formic acid is further mineralized to CO_2 and H_2O by a set of hydroxylation and dehydrogenation reactions (step J).

Figure 7 directly tracks the electron and C equivalents in all the organic intermediates, and it shows the release of N and S as NH_3 and SO_4^{2-} , respectively. SD contains 40 e^- equivalents, 10 equivalents of C, 4 equivalents of N, 1 equivalent of S.

Looking first at electron equivalents in carbon, Steps A and B are mono-oxygenation reactions that require 2 e^- equivalents each, shown as 2H. From step C to step H, transformation of ABS requires 4H for the mono-oxygenations in Steps E and G, but then generates 24H in steps G and H. Therefore, the net yield of electron equivalents from full mineralization of ABS is +20H. However, the pathway must proceed past step G to gain the net yield of e^- equivalents, because the net yield is zero up through step G. Thus, step H must occur in order that ABS transformation generate excess H that can accelerate the initial mono-oxygenation reactions. Mineralization of 2-amino-4,6-dihydropyrimidine (Steps I and J) has a net yield of +8H, but it requires getting past step I.

Mineralization of the C in SD requires that steps H and J occur, since no CO_2 is released in prior steps. 60% of the C originally present in SD is released in the ABS pathway, steps D through H; the remaining 40% are routed through Steps I and J. The TOC results of Figure 2 (60% loss of TOC) suggest that step H occurred, but step J did not occur. This means that the remaining C accumulated as formic acid.

Turning now to the fates of N and S, the 100% releases of S and N (Figure 6) are consistent with full mineralization loss via steps D through H (all of the S and 25% of the N) and that step I occurred (the other 75% of the N). This reinforces that step H occurred, even though the TOC results (Figure 2) suggest that step J did not occur.

In summary, the SD biodegradation pathway, based on a diverse set of the experimental results, explains that the mineralization of ABS (via An) provided internal electron carriers that accelerated the initial mono-oxygenation reactions of SD biodegradation. The faster initial transformation of SD in the P&B experiments (Fig. 1, 2, and 5), as well as with addition of ABS and An (Fig. 5), the 60% mineralization (Fig. 2), and the parallel stoichiometric release of S and N (Fig. 6) support that the ABS and An produced by intimately coupled photolysis accelerated initial mono-oxygenation reactions via the production of intracellular electron carriers at Step H.

4. Conclusions

Our experimental results demonstrate that SD biodegradation and mineralization were accelerated by means of intimately coupled UV photolysis. Photolysis of SD generated intermediates (An and ABS) whose biodegradation generated electron equivalents (H) that stimulated initial SD mono-oxygenation reactions.

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Table and Figure Captions:

Table 1.

Substrates prepared with different component concentration (mg/L) for biodegradation (B) experiments.

Table 2. Concentrations (mg/L) contributed by 20-mg/L (0.076 mM) SD and its photolysis products.

Figure 1. SD removal for P, B, P&B, and control experiment in the ILPBR. The error bars for protocols P, B and P&B came from the average values of the 2 experimental results.

Figure 2. TOC removal percentages for the B and P&B experiments at the end of 4 hours. The P experiment had no TOC removal and is not shown.

Figure 3. Formation of aniline (An), 2-aminopyrimidine (2-AP), *p*-aminobenzenesulfonic acid (ABS), sulfate, and ammonia during SD photolysis. TOC removal was negligible and is not shown.

Figure 4. Proposed SD-photolysis pathway with the number of electron equivalents (e^-) in C shown for each compound. Balanced reactions for each step are as follows, where H stands for one electron equivalent, or $H^+ + e^-$:

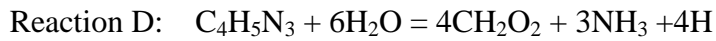
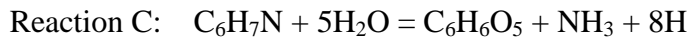
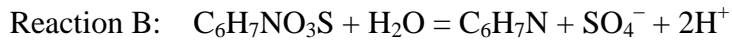
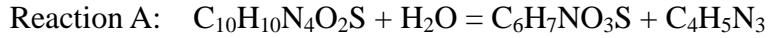


Figure 5. The effect of adding the different photolysis products on SD biodegradation rates. The regression lines are for zero-order kinetics and for data after 10 minutes.

Figure 6. N and S released during SD biodegradation in B and P&B experiments with a starting SD concentration of 10 mg/L. N1 and S1 are N and S releases that correspond to 100% mineralization of SD. N2 and S2 are the measured NH_3^+ -N and SO_4^{2-} -S concentrations during the experiments.

Figure 7. Proposed SD photo-biodegradation pathway. Each organic intermediate is identified by its name, molecular formula, and numbers of electron (e^-) and C equivalents in its organic carbon.

Table 1. Substrates prepared with different component concentration (mg/L) for biodegradation

(B) experiments						
Component \ Group						
	SD(O)	SD(P)	SD+An+2-AP+ABS	SD+An	SD+2-AP	SD+ABS
SD	16	16	16	16	16	16
An	0	1.15 [*]	1.15	1.15	0	0
2-AP	0	3.30 [*]	3.30	0	3.30	0
ABS	0	0.29 [*]	0.29	0	0	0.29

* Produced from UV photolysis

Table 2. Concentrations (mg/L) contributed by 20-mg/L (0.076 mM) SD and its photolysis products.

Compound	Molecular weight	Compound mg/L	Molarity mM	N mgN/L	S mgS/L	TOC mgC/L
Initial Sulfadiazine (SD)	250	18.91	0.076	4.230	2.417	9.064
(C₁₀H₁₀N₄O₂S)^a						
<i>p</i>-aminobenzenesulfonic acid (C₆H₇NO₃S)	173	1.386	0.008	0.112	0.256	0.576
2-aminopyrimidine (C₄H₅N₃)	95	6.513	0.068	2.876	0	3.287
Aniline (C₆H₇N)	93	6.169	0.066	0.927	0	4.769
Sulfate (SO₄²⁻)	96	6.423	0.066	0	2.141	0
<i>2-Hydroxyhexa-2,4-diendioic acid (C₆H₆O₅)^b</i>	<i>158</i>	<i>0.790</i>	<i>0.005</i>	<i>0</i>	<i>0</i>	<i>0.240</i>
Ammonia (NH₃)^c	17	0.085	0.005	0.070	0	0
<i>Formic acid(CH₂O₂)</i>	<i>46</i>	<i>0.920</i>	<i>0.020</i>	<i>0</i>	<i>0</i>	<i>0.240</i>
Ammonia (NH₃)^d	17	0.255	0.015	0.210	0	0
Element sums for products ^e				4.195	2.397	9.112

^a **Boldface** indicates a directly measured concentration.

^b *Italics* indicate that the concentration was computed from the N, S, and C mass balances.

^c This NH₃ comes from reaction C.

^d This NH₃ comes from reaction D.

^e This row is sum of all SD photolysis products.

Figure 1

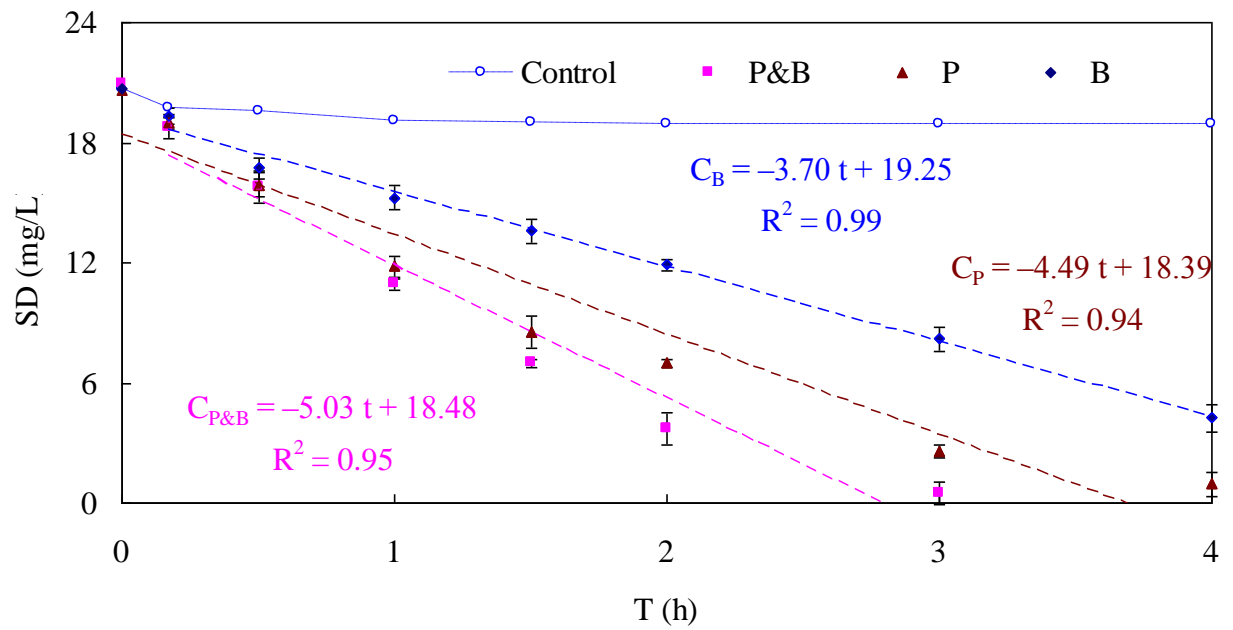


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Figure 2

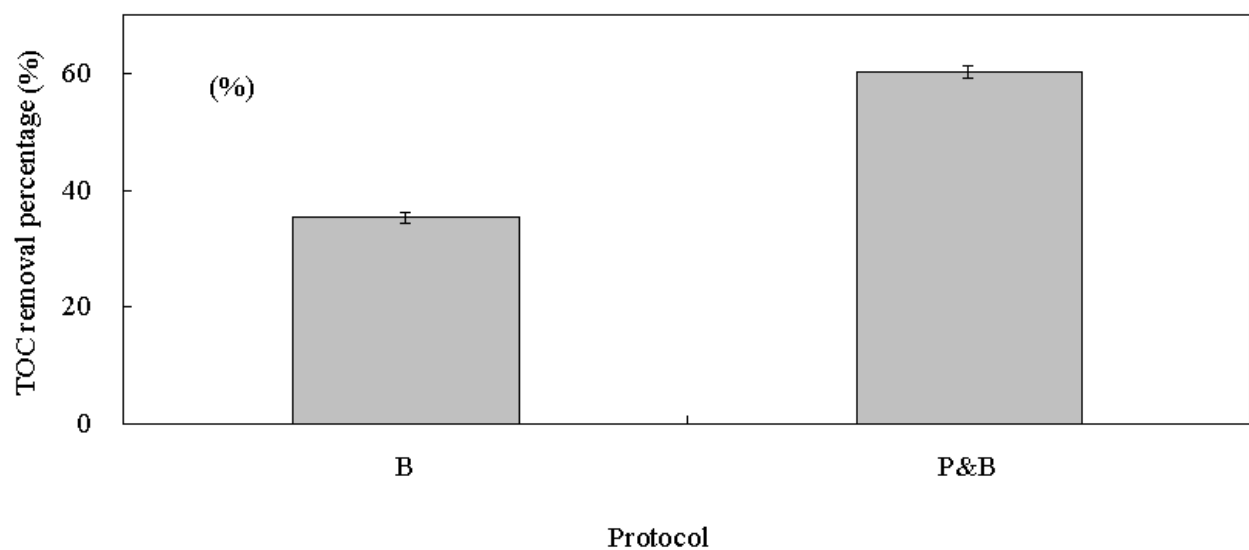


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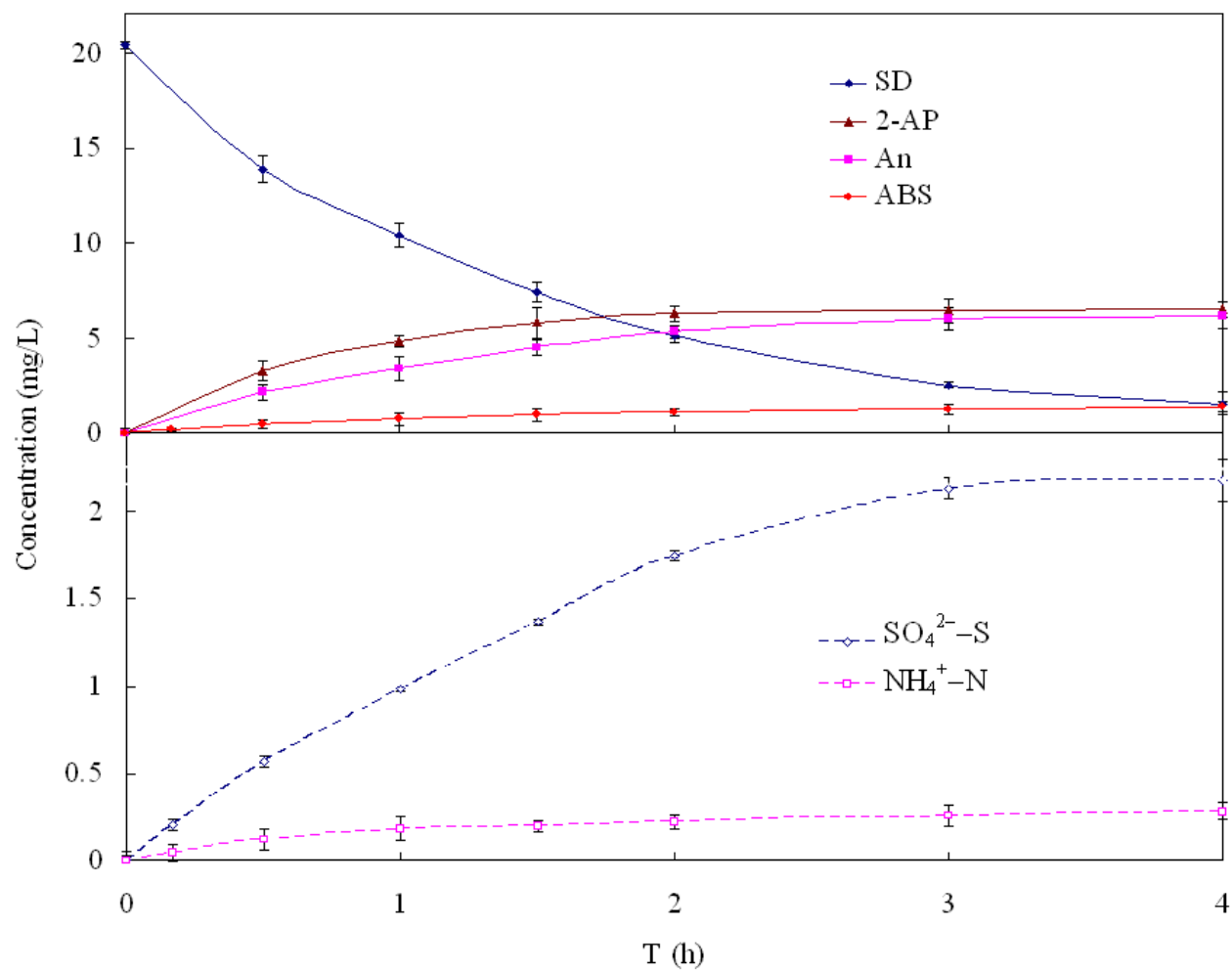


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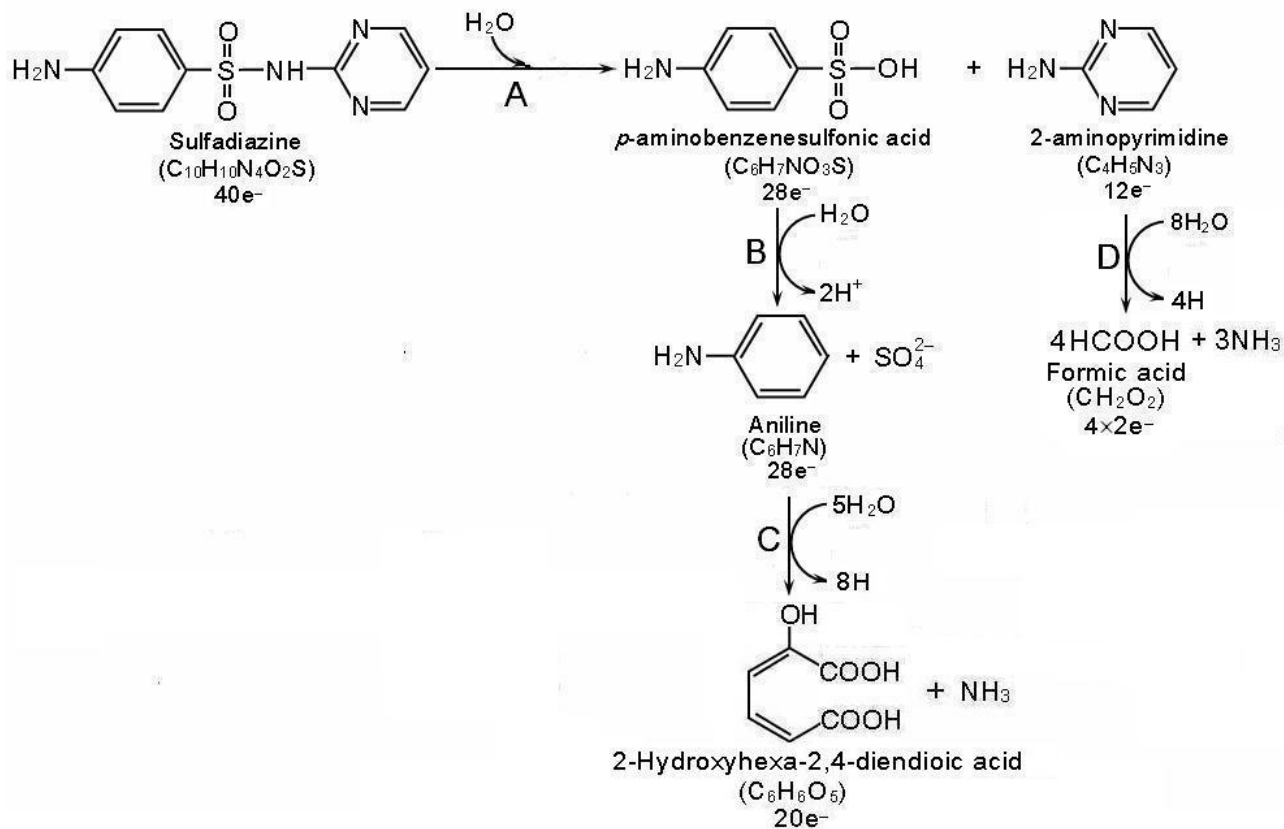


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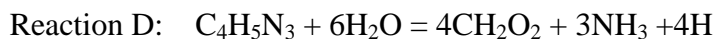
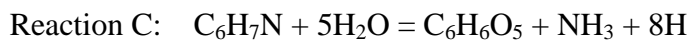
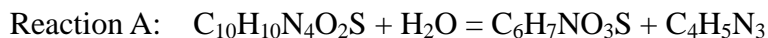


Figure 5

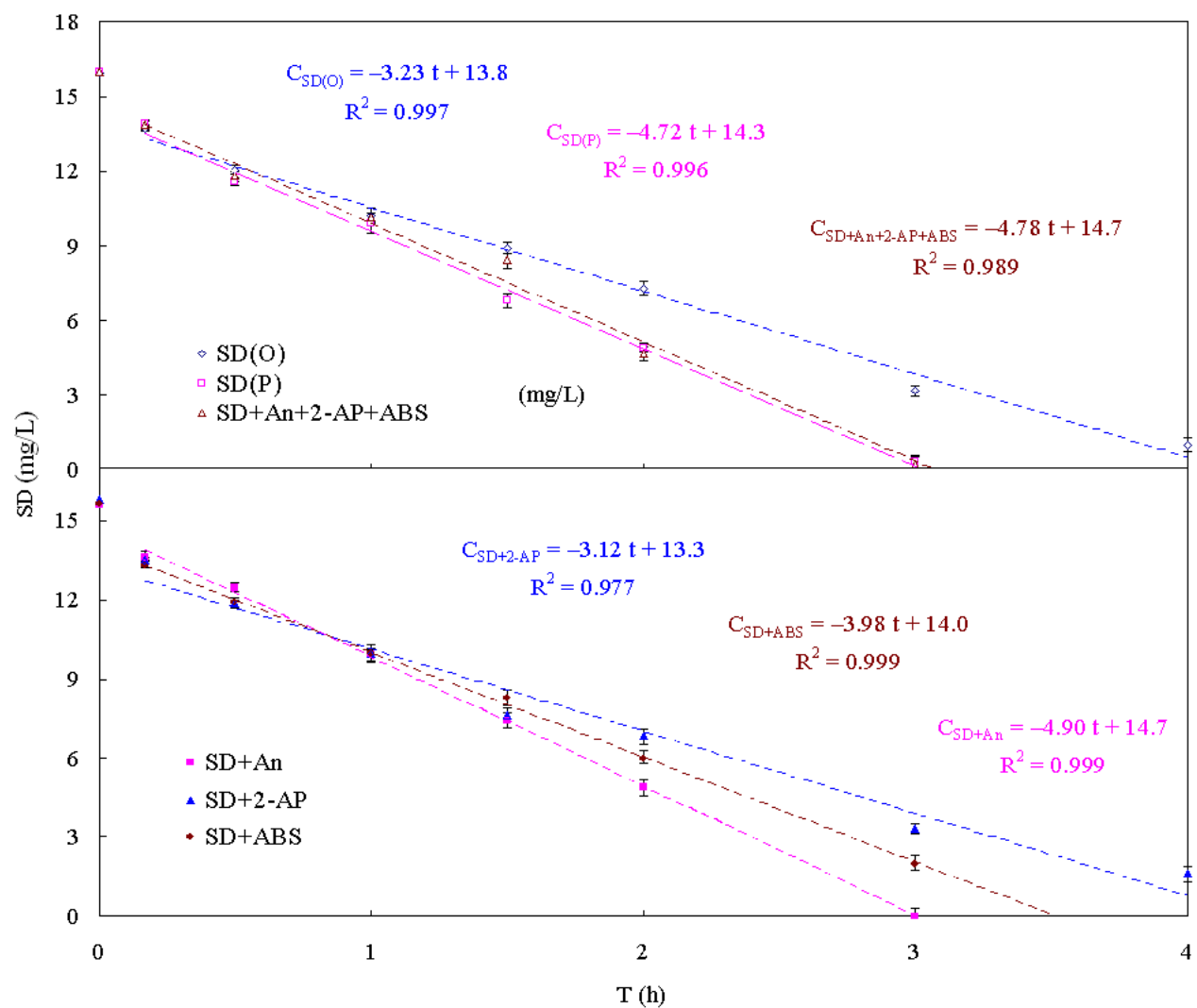


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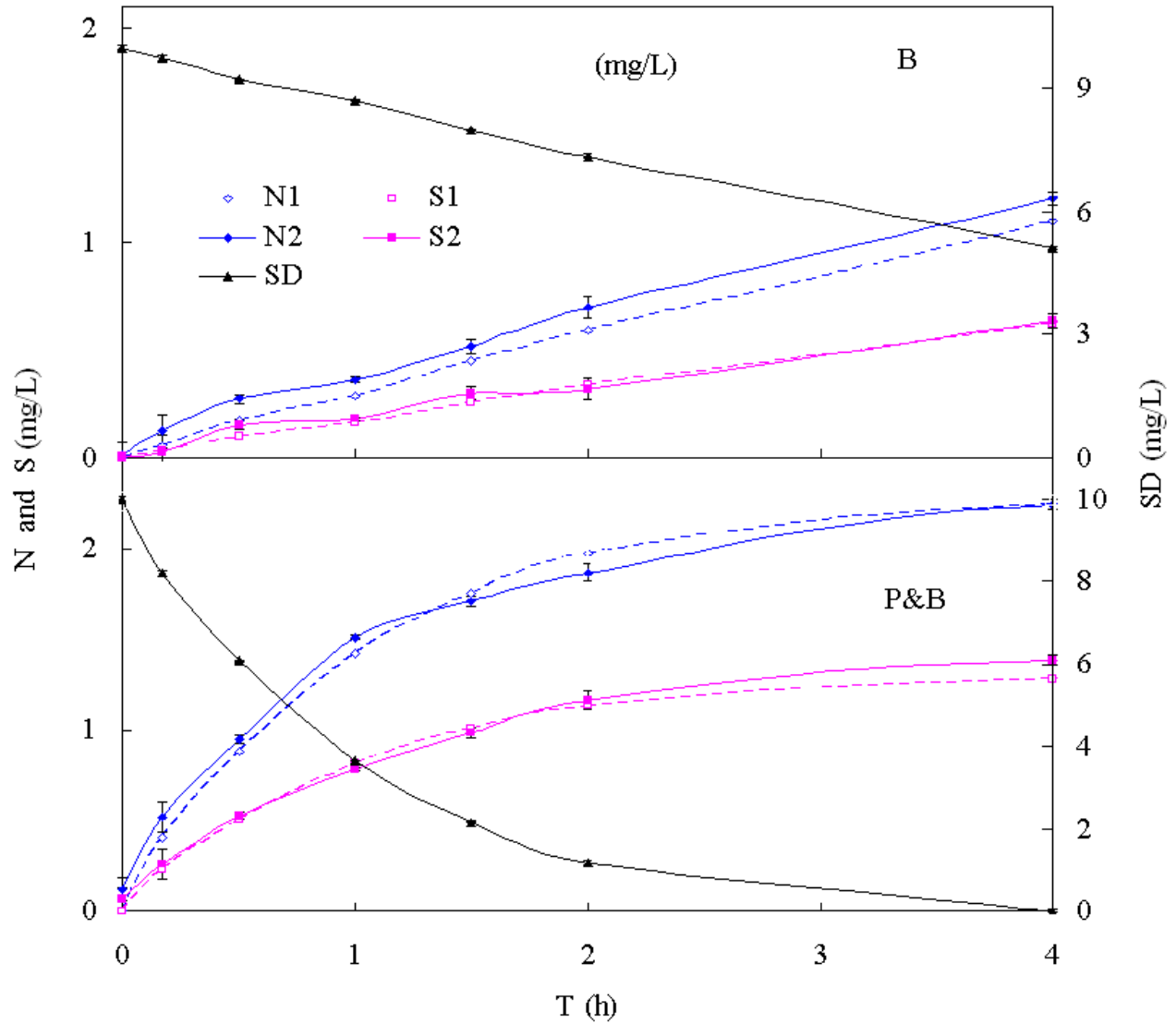


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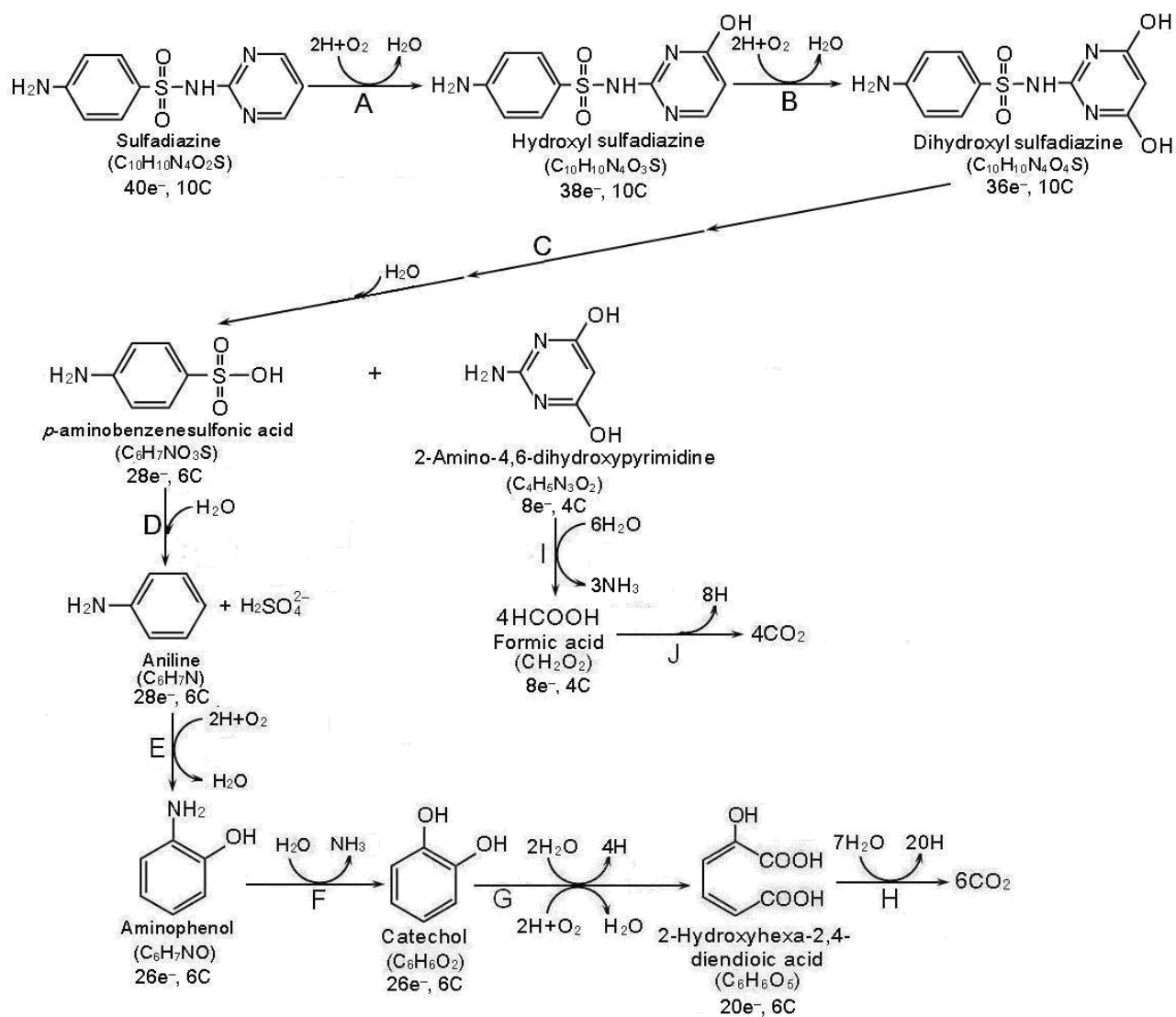


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Cover Letter

May 15, 2014

Dear Editor,

Enclosed please find our manuscript entitled, “**How UV photolysis accelerates the biodegradation and mineralization of sulfadiazine (SD)**” to be considered for publication in *Biodegradation*.

This research explores the mechanism by which UV photolysis accelerates sulfadiazine (SD) biodegradation. We employed an internal loop photobiodegradation reactor (ILPBR) to realize intimately coupled UV photolysis with biodegradation following three protocols: biodegradation alone (B), photolysis alone (P), and photobiodegradation (P&B). The experimental results showed that P&B was superior to B and P in terms of accelerating SD biodegradation and mineralization. We hypothesized that UV photolysis generated biodegradable intermediates whose oxidation accelerated SD biodegradation through the production of intracellular electron carriers needed for initial mono-oxygenation reactions. We evaluated the hypothesis by adding the detected SD photolysis intermediates during biodegradation experiments (no UV photolysis). We found that two of the photolysis intermediates -- *p*-aminobenzenesulfonic acid (ABS) and aniline (An) gave SD removal and mineralization rates similar to intimately coupled photolysis and biodegradation (P&B). We also used mass-balance analyses of B and P&B experiments to develop a biodegradation pathway and demonstrate that ABS and An were fully mineralized to produce the intracellular electron carriers. Thus, we developed multiple lines of evidence support that the mechanism by which intimately coupled photolysis and biodegradation accelerated SD removal and mineralization was through producing co-substrates whose oxidation produced electron equivalents that stimulated the initial mono-oxygenation reactions for SD biodegradation.

I am looking forward to our next correspondence.

Sincerely,

Yongming Zhang, Ph.D., Professor and Chairman

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